

30 Years DGfZ : A Personal View, History and Challenges

Günter Valet

Max-Planck-Institut für Biochemie, Martinsried,
Germany (retired)

30th DGfZ Conference, Berlin, Oct 1-2, 2020

<https://www.classimed.de/valdgz20.pdf>

Major Phases of Early Flow Cytometry Development

- cytometer **construction** (arc lamp, laser, chip based, mass spectrum, imaging in flow...)
- **DNA** (cell cycle, DNA aneuploidy in cancers)
- **antibodies** (immunology: exploratory, clinical)
- **cell functions** like enzymes, pH, Ca²⁺, oxidation, reduction ...)
- **data analysis** (display, information extraction, cluster calculations, predictive medicine by cytomics ...)

History Flow and Image Cytometry

Cellular Pathology

R.Virchow Berlin 1858
cell staining, microscopy (1)

K.v.Rokitansky Vienna
macroscopic pathology

L.Koss Vienna, NY 1961
histo/cell pathology
flow cytometry (7,8)

G.Wied Prague, Munich
1968, **TICAS image analysis system** (5,6)

M.Melamed NY 1963
cyto/histopathology
flow cytometry (10)

Z.Darzynkiewicz
Warsaw, NY 1969
cell biology by
flow cytometry (11)

Cytometry

TO Caspersson
Stockholm 1934

- DNA, protein by
light absorption
- chromosome
banding (2)

B.Mayall LLL 1965
- CYDAC DNA
chromosome analysis
- editor Cytometry (12)

L.Kamentsky
NY 1965
- flow cytometry by
light absorption (9)
- computerized data
evaluation & display
Cambridge, MS 1991
- laser scanning
cytometer (LSC) (30)

Flow Cytometry

W.Coulter
Hialeah, 1949/56

**electronic blood
cell counter** (3/4)

CC Lushbaugh
Los Alamos 1962
biological/right
skew (14/15)

**L.Hallermann, R.Thom,
H.Gerhartz**
Berlin 1964 **flow cytometry
by fluorescence** (31), blood
leukocytes (AO)

M.Fulwyler
Los Alamos 1965
artificial right skew
cell sorter (16)

M.van Dilla
Los Alamos 1968
flow cytometry (19) by
fluorescence (AO,FDA)

L.Herzenberg
Stanford 1969
fluorescence activated
cell sorter (23)

W.Göhde
Münster 1968
- first commercial
fluorescence
flow cytometer
(Phywe ICP11)
- DNA aneuploidy
as tumor marker
(EB) (20-22)

G.Ruhenstroth-Bauer, O.Zang
Munich 1960
artificial/right skew of Coulter cell
volume distribution curves (13,17),
<https://www.classimed.de/martins1.html>

R.Thom, V.Kachel, G.Valet
Munich/Martinsried 1969/79
- *artificial* right skew
- central beam focusing (18)
- fast imaging in flow (24)

G.Valet
Munich/Martinsried 1972/01
- cytometry bioinformatics (25)
- predictive medicine
by cytomics 1987-2001 (26-28)
- cell function in disease (29)

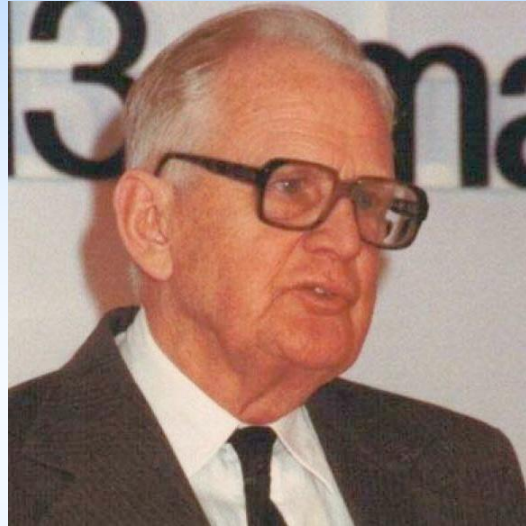
- **European background**
- **US background**
- **innovation**

- references at: <https://www.classimed.de/martins2.html>

Fathers of Cytometry



Rudolf Virchow
(1821-1902)
-cellular pathology



Torbjörn Caspersson
(1910-1997)
-cell DNA determination
by *image* cytometry
-chromosome banding

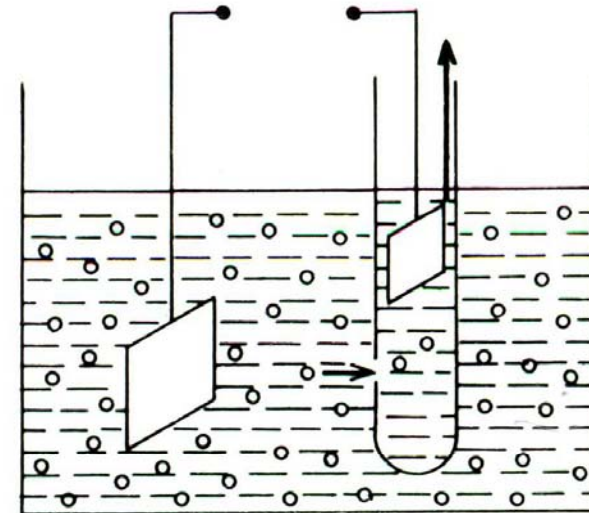


Wallace Coulter
(1913-1998)
-Coulter counter
-*flow cytometry*

Coulter Cell Counting and Sizing



**Coulter model A
counter 1956**



$I = \text{konst.}$

$$\Delta u = \Delta R \cdot I$$

$\Delta R \sim \text{Volumen des Partikels}$

$\Delta u \sim \text{Volumen des Partikels}$

Coulter Cell Counting and Sizing

- **undisputed** counting (Ruhenstroth, Zang 1960)
- *right skew* of Coulter volume distribution curves:
real or artefact?

- ***real***: C.Lusbaugh 1962 (reticulocytes)
- ***artefact***:
 - M.Fulwyler 1965: *cell sorter* but *no* explanation
 - R.Thom, V.Kachel 1970: central beam *hydrodynamic focusing, fast imaging* in flow, explanation of right skew

Electronic Measurement of Cellular Volumes. II. Frequency Distribution of Erythrocyte Volumes

By C. C. LUSHBAUGH, N. J. BASMANN AND B. GLASCOCK

BLOOD, VOL. 20, No. 2 (AUGUST), 1962

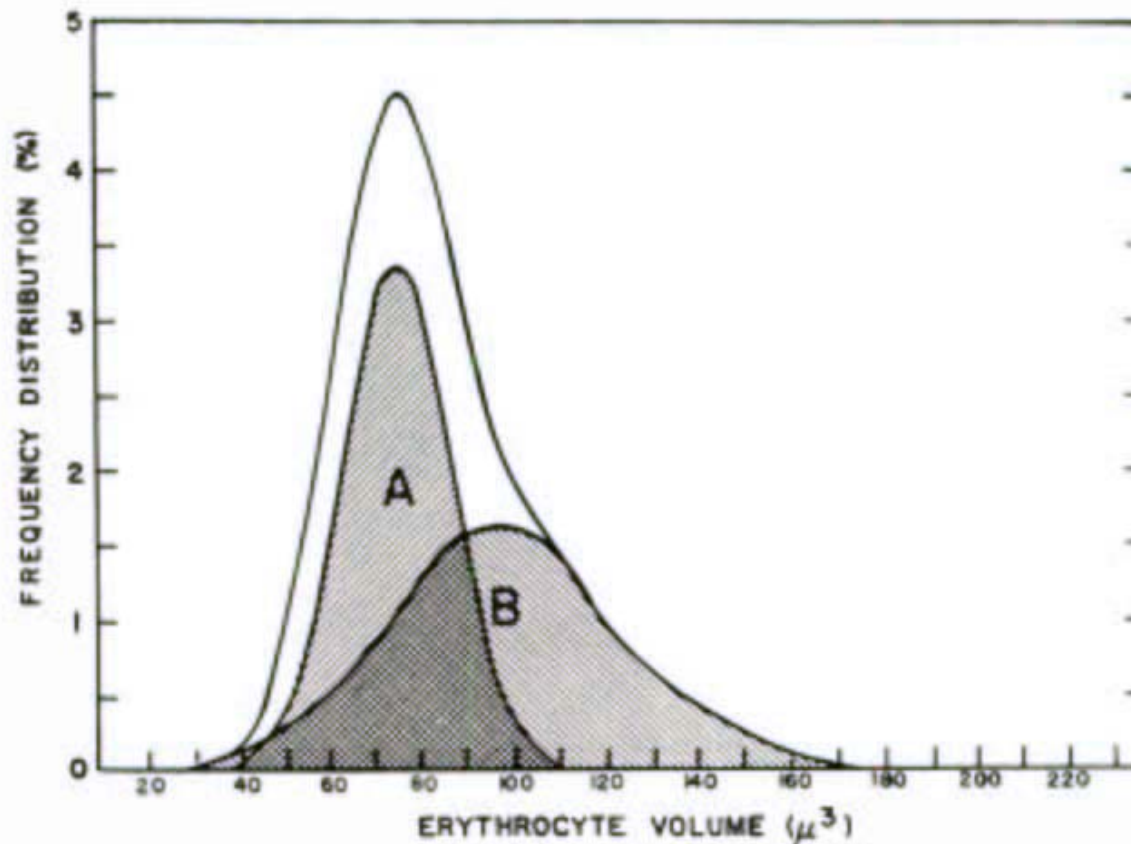
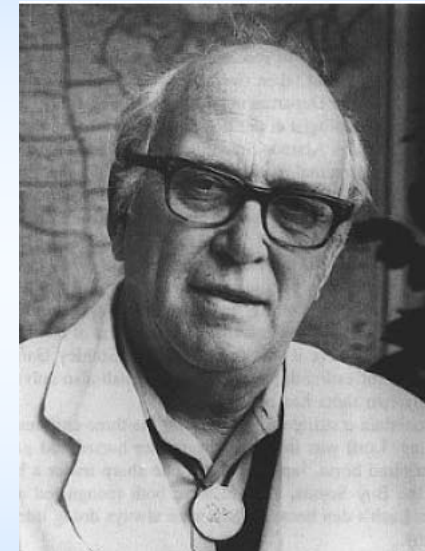


Fig. 2.—Frequency distribution curve of erythrocyte volumes of human blood and its representation by two cell populations (A and B), each with a normal Gaussian distribution.



Clarence C. Lusbaugh
(1916-2000)

Electronic separation of biological cells by volume
MJ Fulwyler
Science 150:910-911(1965)



Mac Fulwyler

Cell Sorter

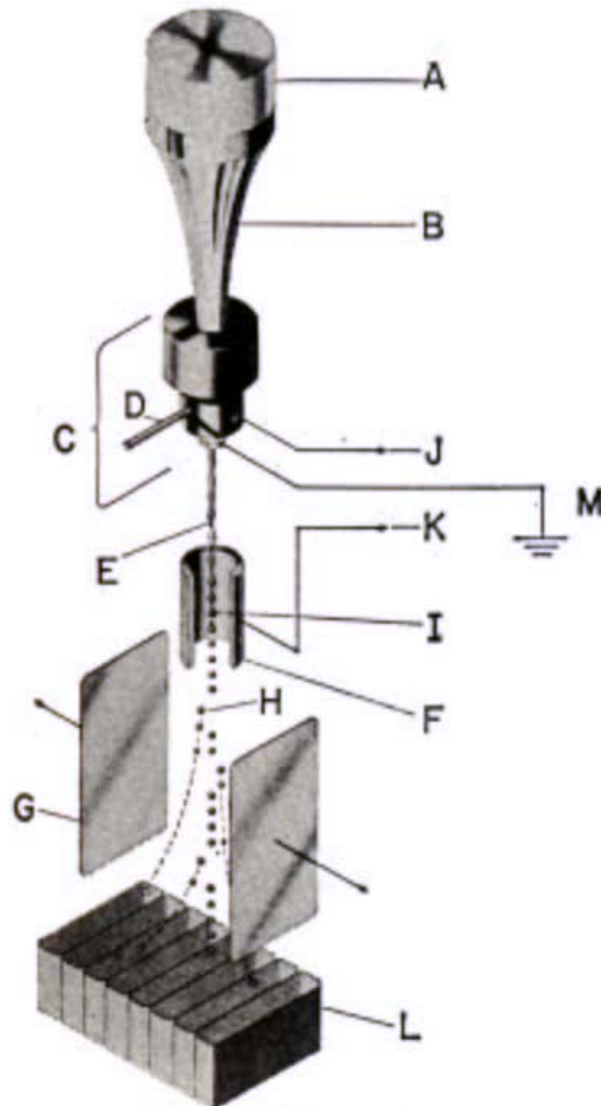


Fig. 1. Cell separator.

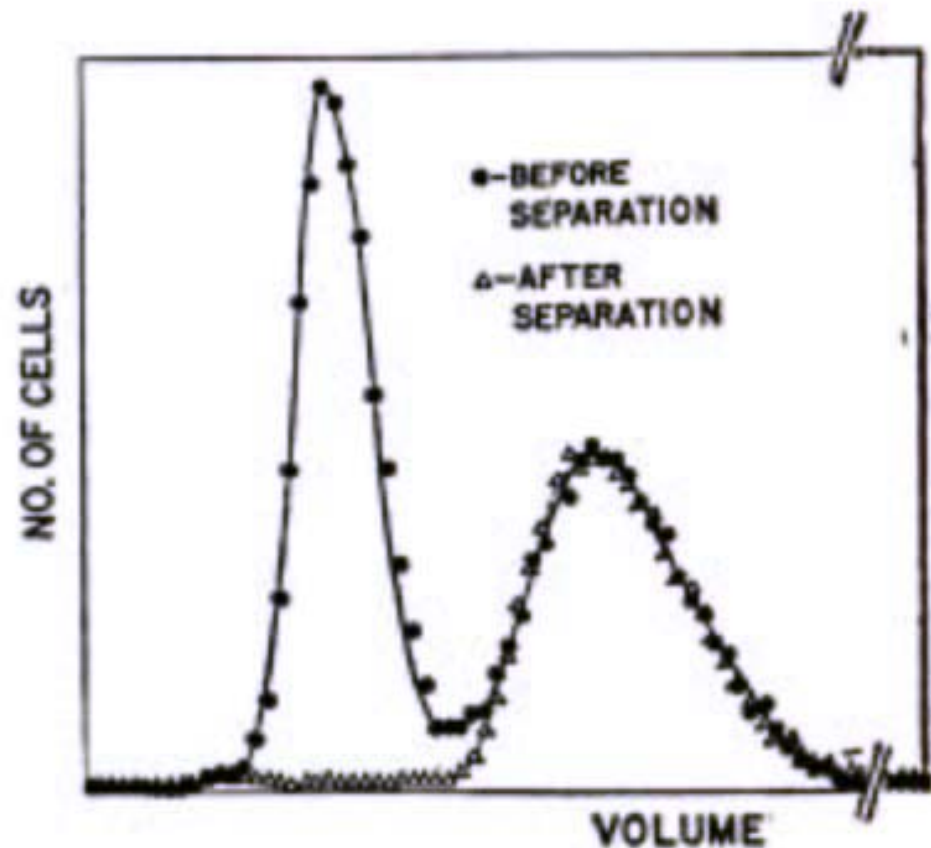


Fig. 2. Distribution by volume of mouse and human erythrocytes before and after separation.

KURZE MITTEILUNG

*Aus der Med. Klinik und Poliklinik der Freien Universität Berlin im Städt. Krankenhaus Westend,
und dem Max-Planck-Institut für Biochemie München*

Fortschritte für die elektronische Größenbestimmung von Blutkörperchen

Von R. Thom und V. Kachel



Reinhard Thom Volker Kachel

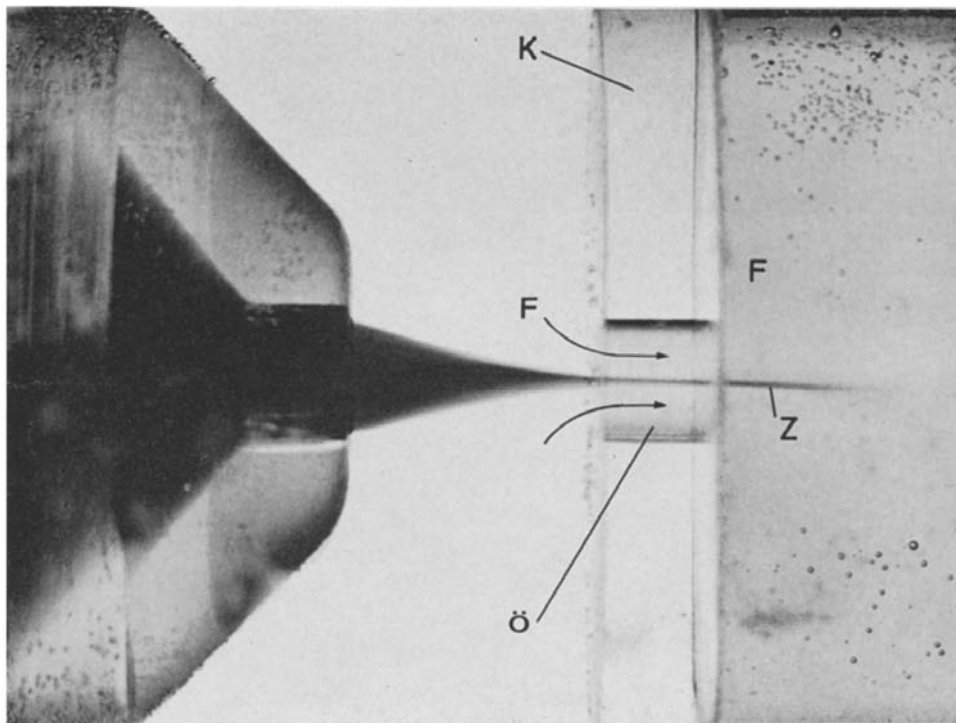


Abb. 1: Meßanordnung K = Kapillarwand; Z = Zentralstrahl, F = Ummantelnde Flüssigkeit; ö = Meßöffnung.

50

R. Thom und V. Kachel

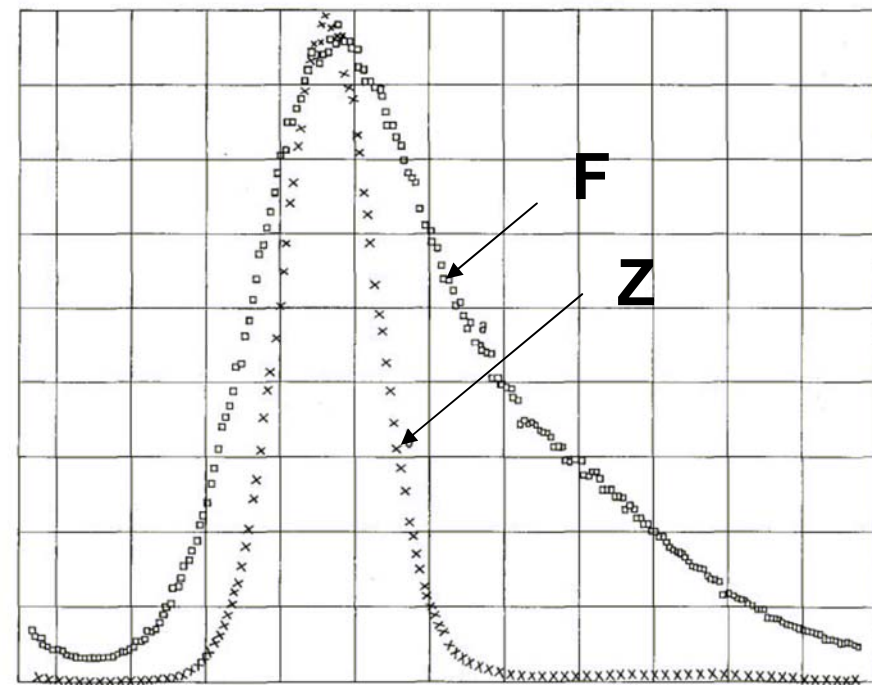


Abb. 3: Volumensverteilungskurven von nativen Erythrozyten. Messung in einer Coulter-Kapillare (100 μ) (a) und in der Zentralstrahlkapillare (b).

Fast Imaging in Flow (1979)

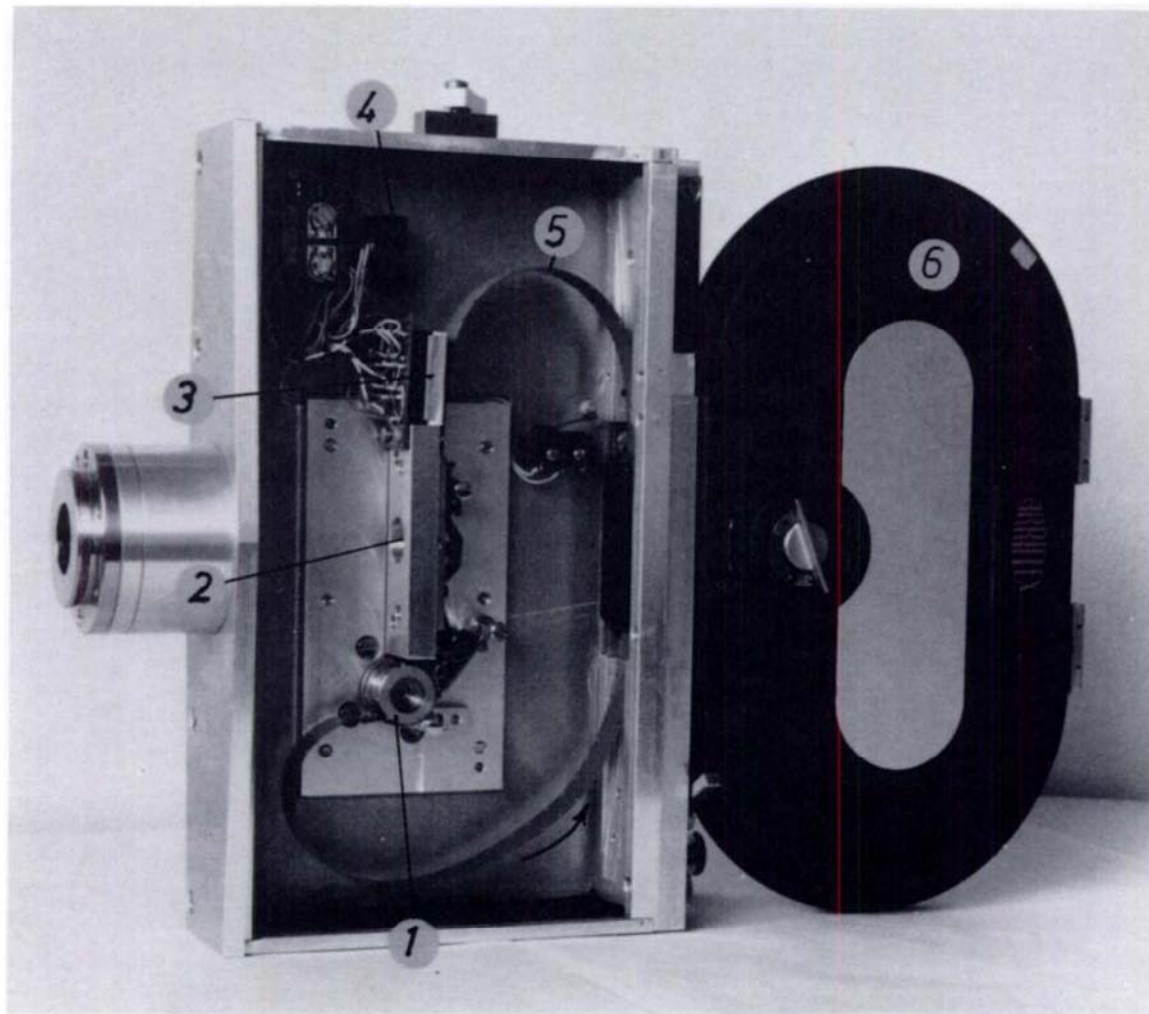


FIG. 4. The camera with Arriflex cassette. 1, sprocket, driven by the step motor; 2, picture gate; 3, three digit hexadecimal film marker; 4, infrared light barrier for controlling the film loop; 5, film loop; 6, Arriflex cassette; the direction of the film motion is indicated by the arrow.

V.Kachel, G.Benker, K.Lichtnau, G.Valet, E.Glossner. J.Histochem. 27:335-341 (1979)

Fast Imaging in Flow (1979)

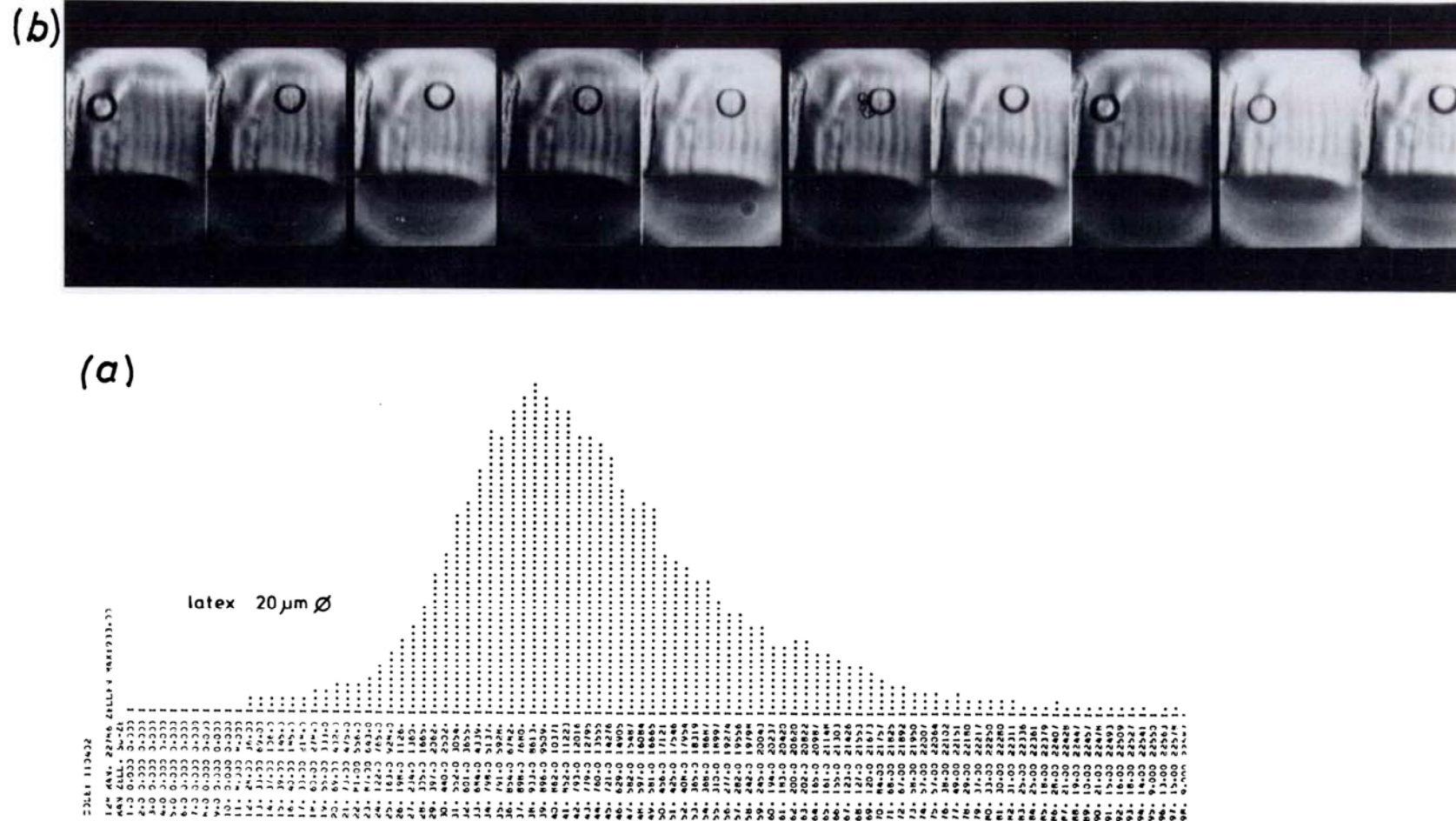


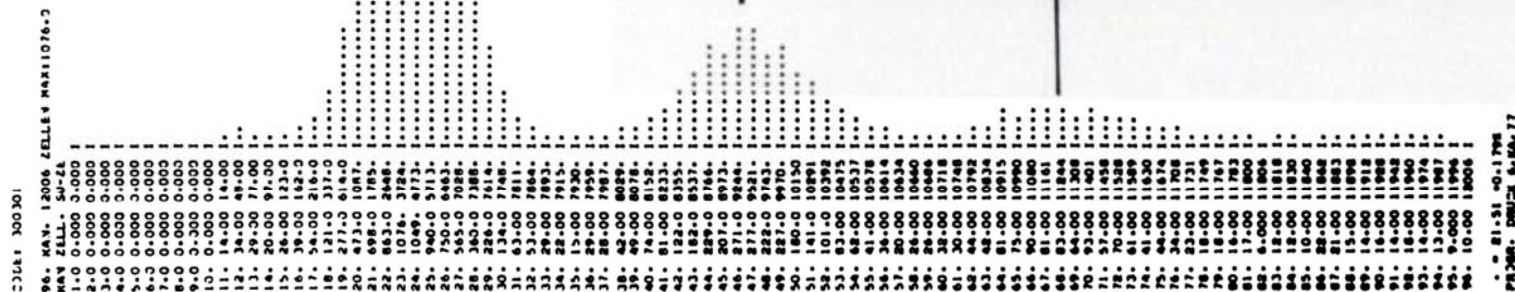
FIG. 7. (a), Volume distribution curve of 20 μm mean diameter latex particles measured with the imaging chamber. Channel width 120 μm . (b), Sequence of particles selected from the right slope of the distribution curve (large particles). Flow direction from bottom to top.

V.Kachel, G.Benker, K.Lichtnau, G.Valet, E.Glossner. J.Histochem. 27:335-341 (1979)

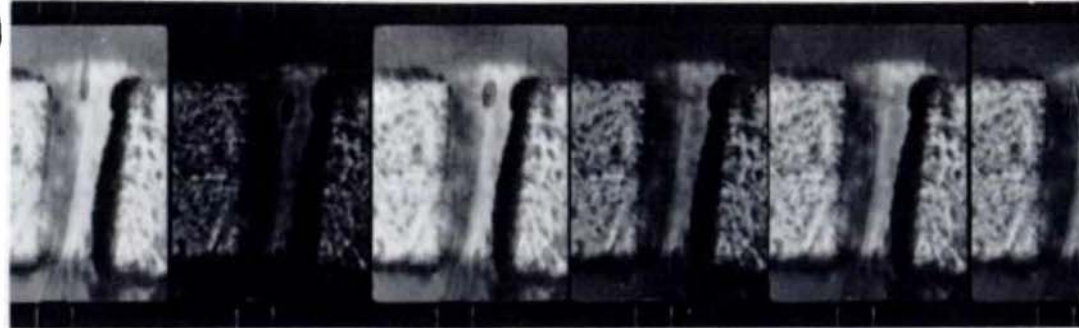
Fast Imaging in Flow (1979)

human erythrocyte
doublets and triplets

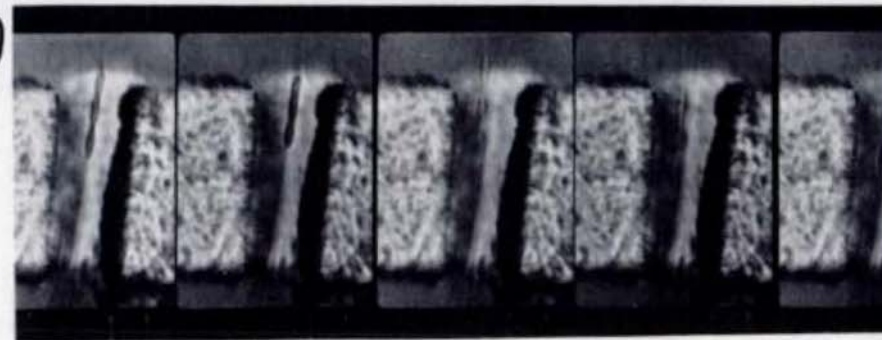
(a)



(b)

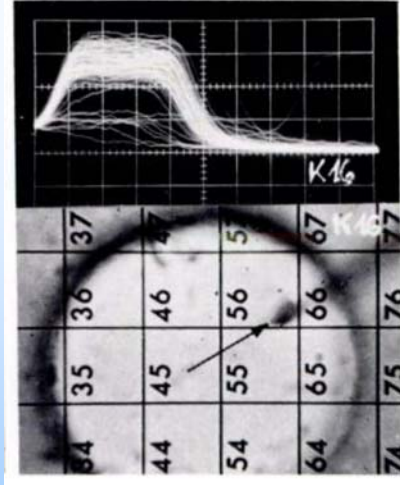
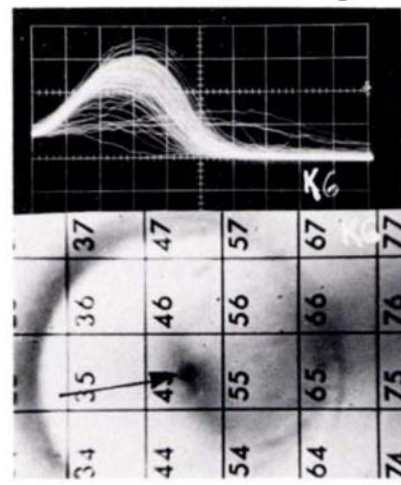
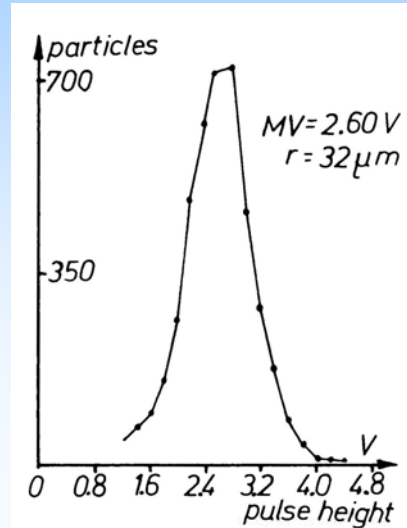
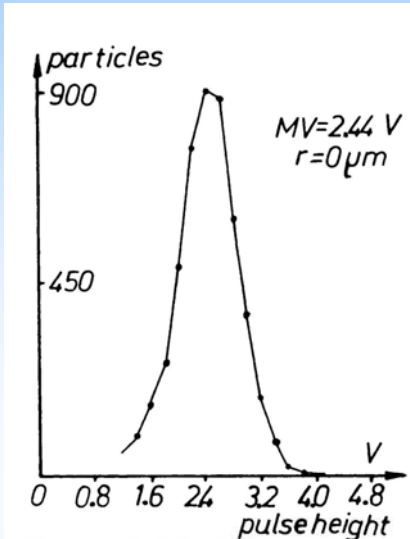
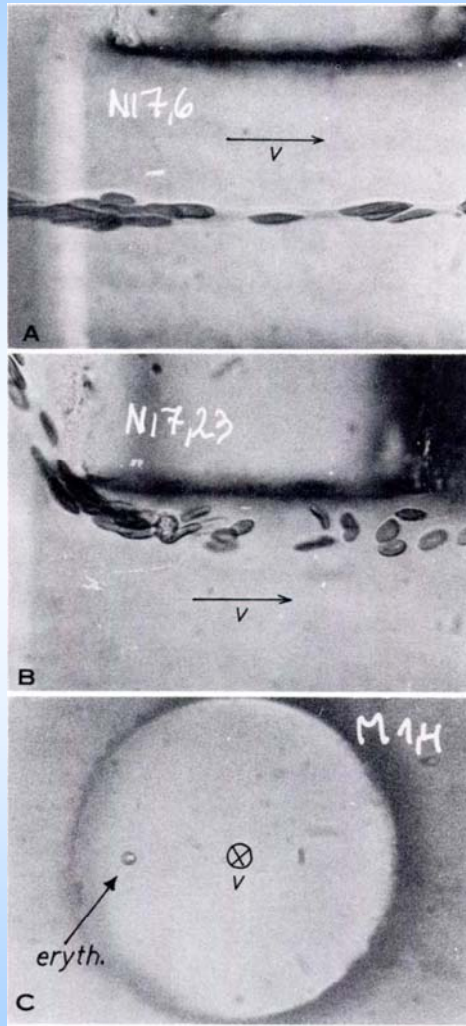


(c)



V.Kachel, G.Benker, K.Lichtnau, G.Valet, E.Glossner. J.Histochem. 27:335-341 (1979)

Fast Imaging in Flow (1979)



- hydrodynamic focusing
- particles remain in *constant* electric field
- result: *symmetric* volume distribution curves (right)
- elongation of native erythrocytes to cigars in the orifice center (left a, c) but tumble over the orifice edges (b)

V.Kachel J.Histochem.Cytochem 24:211-230 (1976)

Flow Cytometer Development in Germany

- **Five** *commercial* flow cytometers were developed in German *scientific institutions* (1969-84).
- **Only** Partec (Münster), since 2013 as Sysmex-Partec (Görlitz) survived

Phywe (Göttingen) ICP-11 (1969)



Wolfgang Göhde,
Universität Münster

- first commercial fluorescence flow cytometer
- acquired by Ortho Diagnostics (Raritan, NJ)
- market removal

High pressure mercury arc lamp, 2 fluorescence channels

Continued Activities with **Partec** (1985)

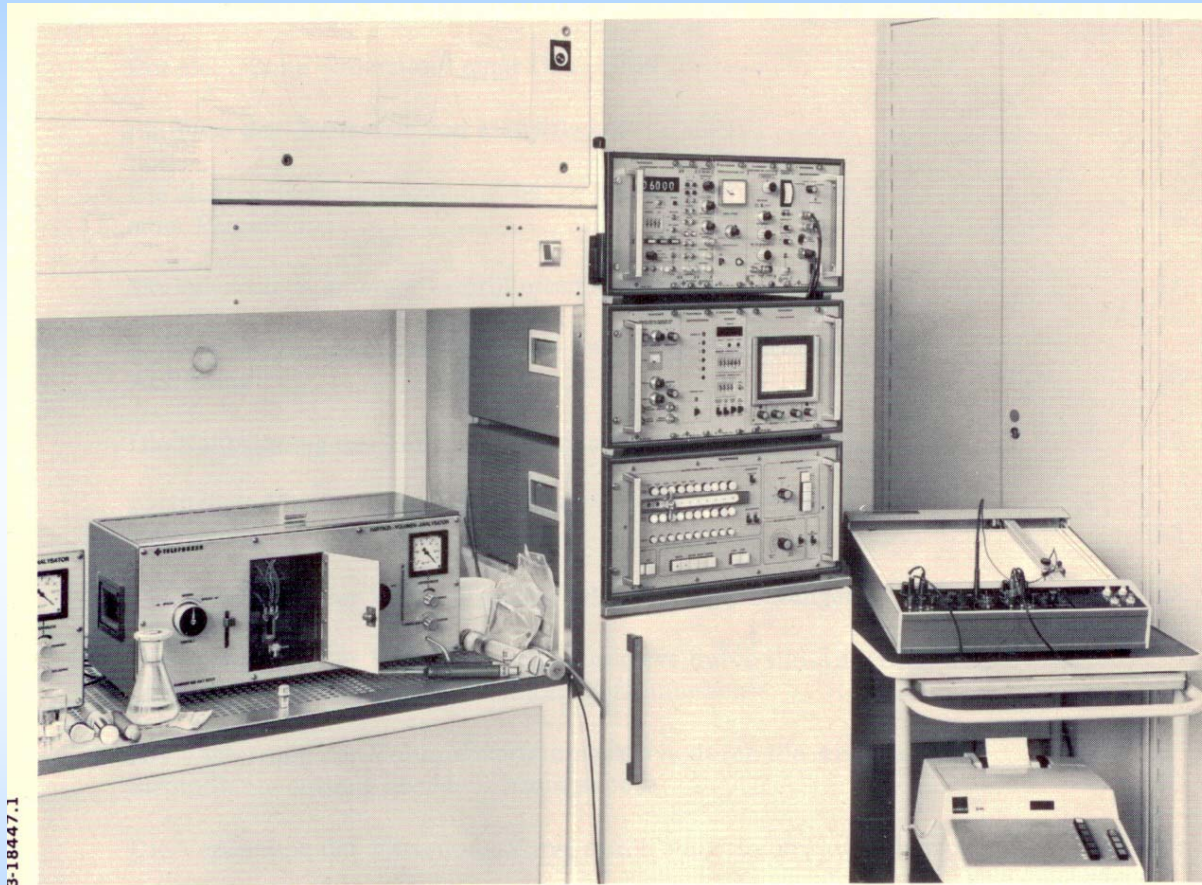


PAS-III flow cytometer

- Ortho had also acquired **all** Phywe flow cytometry patents, being internationally valid until 1988, with the **exception** of *Switzerland* where *Wolfgang Göhde* continued flow cytometer development (PAS-I, PAS-II) with his own **Partec** company, moving upon patent expiration to *Münster* (PAS-III) after 1988 and in part to *Görlitz* following the German reunification in 1989.
- **Partec** was acquired 2013 by Sysmex (Kobe, JP) and continues activities as **Sysmex-Partec**

Hydrodynamically focused Coulter cell volume, laser FSC, SSC,
2 or 3 fluorescence channels, piezo driven fluidic cell sorting chamber

AEG-Telefunken (Berlin) MPV-1 (1982)



3-18447.1

Vollständige Meßanlage zur Partikelvolumen-Analyse

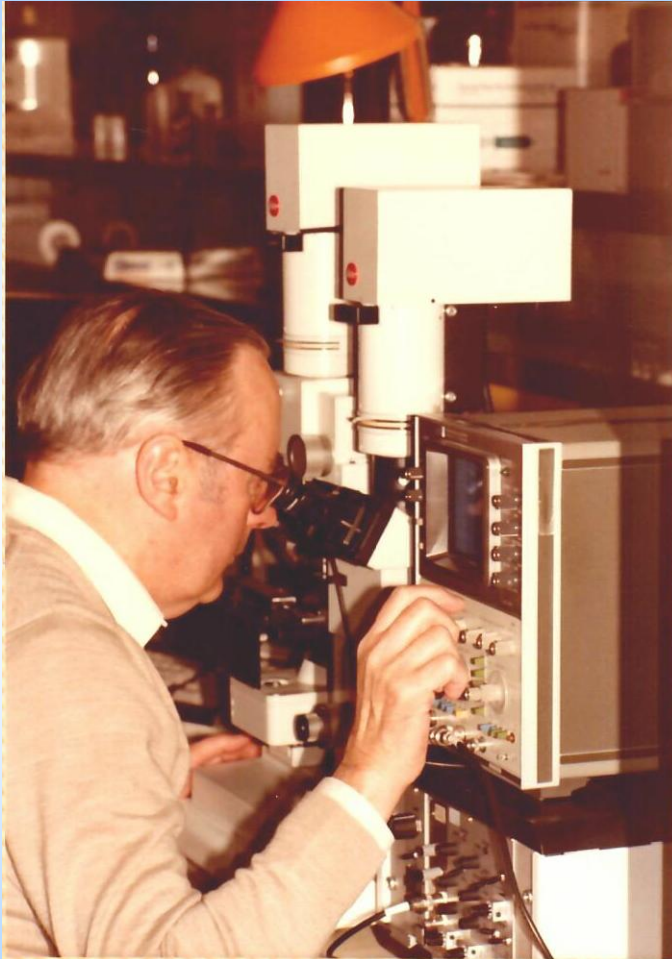
Reinhard Thom
Westendkrankenhaus
Berlin sowie

A.Hampe, G.Sauerbrey
Physikal. Techn. Bundesanstalt
Braunschweig

- acquired by Coulter Electronics (Hialeah, FL)
- **market removal**

1 channel hydrodynamically focused Coulter cell volume instrument

Leitz (Wetzlar) MPV-Compact Flow Cytometer (1982)

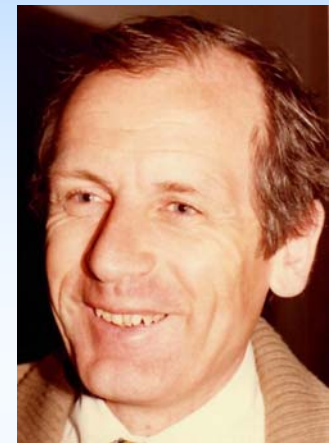
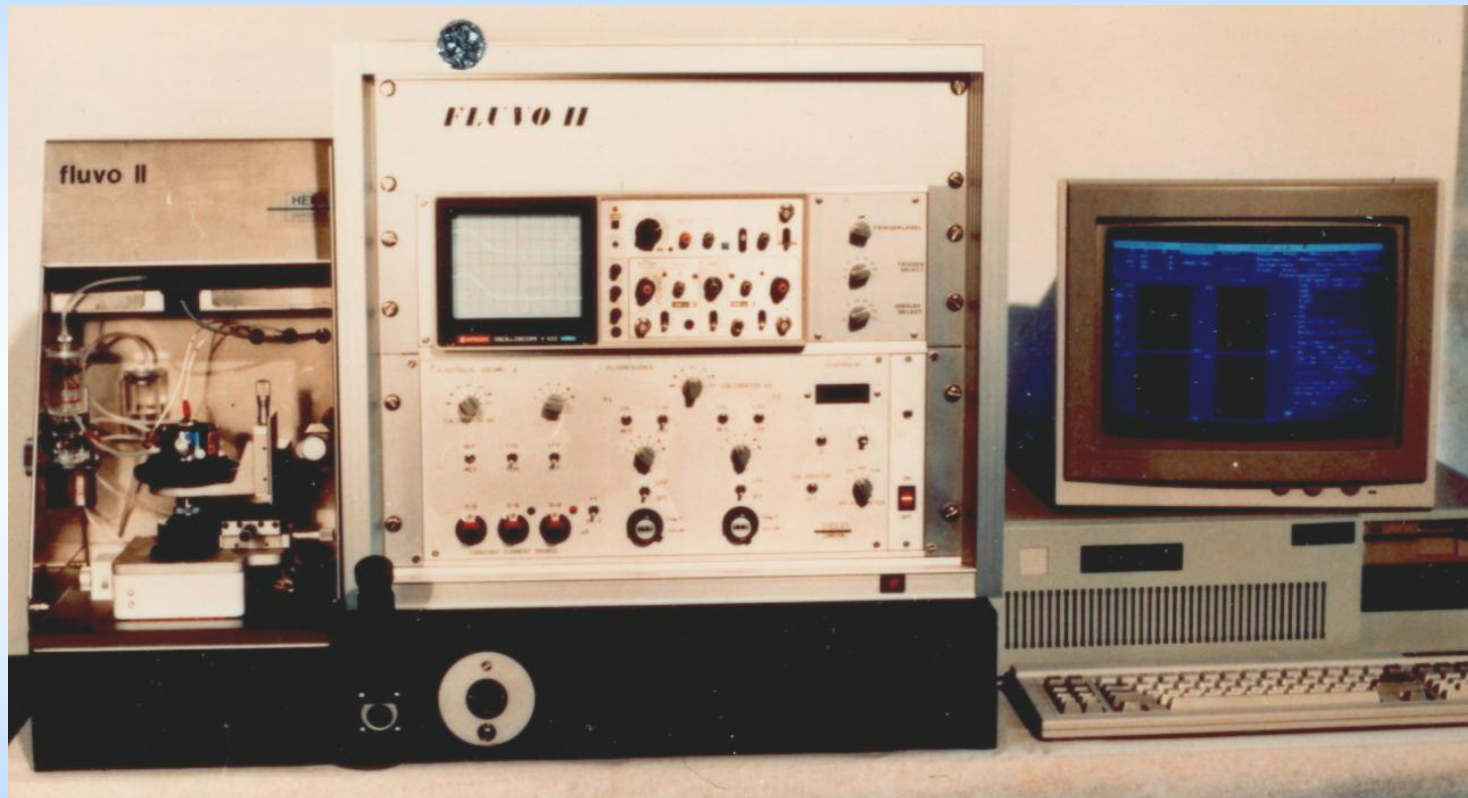


Harold Steen
Oslo (Norway)

This flow cytometer was particularly **conceived for** small particle analysis, like bacteria, cell nuclei, or mitochondria

High pressure mercury arc lamp, 2 light scatter + 2 fluorescence channels

HEKA (Lambrecht/Pfalz) FLUVO-Metricell (1983)



Volker Kachel
Max-Planck Inst.
für Biochemie
Martinsried

Hydrodynamically focused Coulter cell volume with high pressure mercury arc lamp and 2 fluorescence channels

Kratel Instruments (Böblingen) Flow Cytometer (1984)



Wolfgang Eisert Wolfgang Beisker
Gesellschaft für Strahlenforschung (GSF,
Hannover)

- The instrument was particularly conceived for precise **particle length measurements** and small particle analysis (clean water in clean room environments)

Argon laser excitation, 2 fluorescences

Cytometry Organizations

- Flow and image cytometers, software, stains and applications were developed in the 1970s by around **150-200 scientists** worldwide mostly from US, Germany and Norway.
- The **Phywe ICP-11** flow cytometer introduction stimulated preferentially *clinical*/tumor cell **DNA aneuploidy** measurements, resulting in a **boost** of publications (**60** mostly German versus **44** searchable US publications between 1969-76).
- Duplicate** result presentations at European and US cytometry meetings prompted 1978 the **foundation** of the **Society for Analytical Cytology** (SAC) at the **American Engineering Foundation** meeting in **Schloss Elmau** near *Mittenwald* (Germany), conceived by *Sandford Cole* and organized by *Klaus Goerttler* Deutsches Krebsforschungszentrum Heidelberg (DKFZ)

Foundation of the Society for Analytical Cytology

European effort

motivator: W.Göhde

Phywe ICP-11/22 mercury arc cytometer

1972 Heidelberg (M.Andreeff)

1973 Nijmegen (C.Haanen)

1975 Münster (W.Göhde)

1977 Vienna (D.Lutz)

1979 Voss (O.Laerum)

1980 Rome (F.Mauro)

American Engineering Foundation

conference organizer: Sanford S.Cole

BioPhysics/Ortho/Coulter/BD laser cytometers


1972 Saxton River, VE

1973 Asilomar, CA

1975 Asilomar, CA

1976 Pensacola, FL

1978 Schloß Elmau/Mittenwald



foundation of the **Society for
Analytical Cytology (SAC)**

& journal **Cytometry**

in Elmau 1978, **first meeting** 1981

in Wentworth by the Sea (NH)

SAC Founding Committee at Schloss Elmau, Germany 1978



back: Myron Melamed, Scott Cram, Sandford Cole, Mort Mendelsohn, Hans Aus, Klaus Goerttler, Jim Tucker, Paul Mullaney, Volker Kachel, unidentified, Brian Mayall, Mac Fulwyler

front: Leon Wheelless, Ted Young, Marvin vanDilla, Dennis Rutovitz, Tom Jovin

Centralized SAC or International Cytometry Network ?

- Fast flow cytometer development, high US company investments and instrument sales **increased** SAC US membership within a few years to around 600 with SAC and later ISAC understanding itself as an international scientific **marketing** organization for US cytometers with **affiliated** national societies.
- **European** cytometry scientists strengthened their position by founding
 - **ESACP** (1986) and the **ACP** journal (today **Cellular Oncology**, **IF 4,19**)
 - national cytometry societies like **DGfZ** (1989)
 - by setting up the worldwide first *cytometry society* Internet servers (**ESACP**, **DGfZ** 1994) and
 - by organizing the first European flow cytometry course for scientists (**Martinsried** 1985-1993) with initially *only* European cytometers
- Today, the national cytometry societies are **associated** with ISAC as an **international** cytometry network

1.-6.Martinsried Flow Cytometry Courses (1985-1993) for >200 scientists

P R O G R A M

1.MARTINSRIED FLOW CYTOMETRY COURSE

4/15-4/19/1985

photographs by *Rudi Kratel*:
<https://www.classimed.de/mk85phot.pdf>

Mildred-Scheel-Laboratory for Cancer Cell Research
Max-Planck-Institute for Biochemistry
8033-Martinsried near Munich





1986: foundation of

European Society for Analytical
Cellular Pathology (ESACP) by:

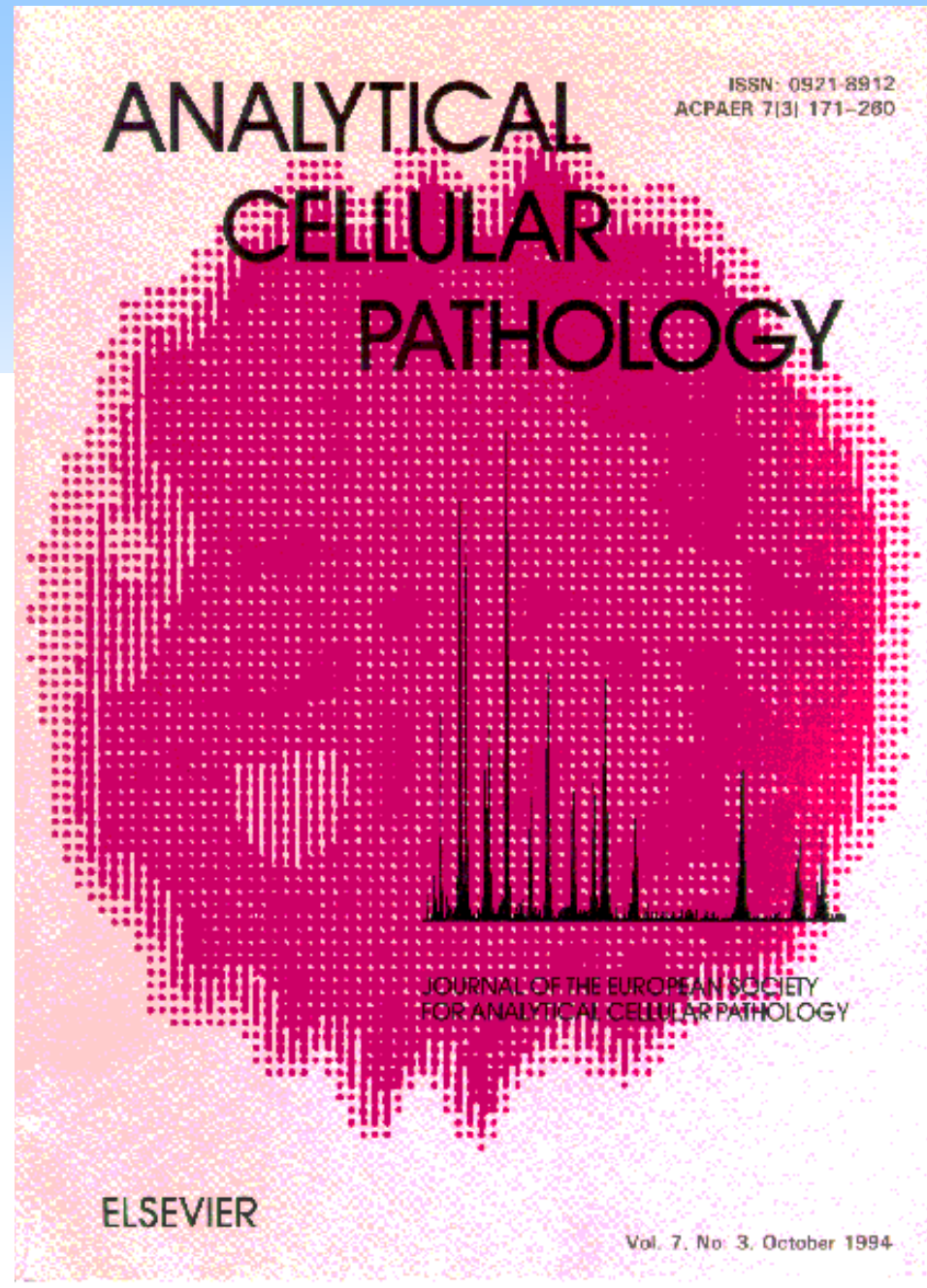
G.Auer, P.Benedetti, G.Brugal, G.Burger,
Y.Collan, C.J.Cornelisse, O.A.N.Husain,
A.Reith, F.Rilke, D.Rutovitz, E.Sprenger,
B.Stenkvis, G.Valet, P.Viallet, L.Vindelov,
G.P.Vooijs.

with the **ACP** journal

G.Burger, G.Valet, P.Vooijs, G.Brugal

continued since 2003 as:

**International Society for Cellular
Oncology (ISCO)** with the **Cellular
Oncology** journal (IF 4.191 in 2019)



DGfZ Foundation 1989

foundation committee

K.Goerttler
C.Cornelisse
G.Feichter
W.Göhde
H.Hoehn
F.Otto
A.Radbruch
G.Valet

membership cohesion

DGfZ treasurer 1990/2010

P.Schwarzmann

DGfZ presidents

90/92 K.Goerttler
92/94 G.Valet
94/96 A.Radbruch
96/98 J.Hemmer
98/00 M.Nüsse
00/02 R.Knüchel
02/04 M.Stöhr
04/06 A.Tarnok
06/08 G.Brockhoff
08/10 S.Müller
10/12 E.Endl

meetings

Heidelberg 1988-2004

K.Goerttler
M.Stöhr
K.Hutter
H.zur Hausen

Leipzig

A.Tarnok 2005/06
S.Müller 2009/10

Regensburg 2007

G.Brockhoff

Bremen 2008

G.Rothe

Future

- **Basic research**: more parameters, miniaturization, mass cytometry, multidimensional data analysis ...
- **Medicine**: exploitation of the **systemic** potential of cytometry (**system cytometry***) like predictive medicine by **cytomics**
- **Concept**: diseases are caused by molecular changes in cells and tissues (*cells know it first*). Molecular cell data patterns permit **individualized** disease course predictions

*) <http://www.cyto.purdue.edu/cdroms/cyto3/8/valet/keyvirt1.htm> (1997)

Basic Research

Dimension reduced multiparameter cluster display in vaccine development

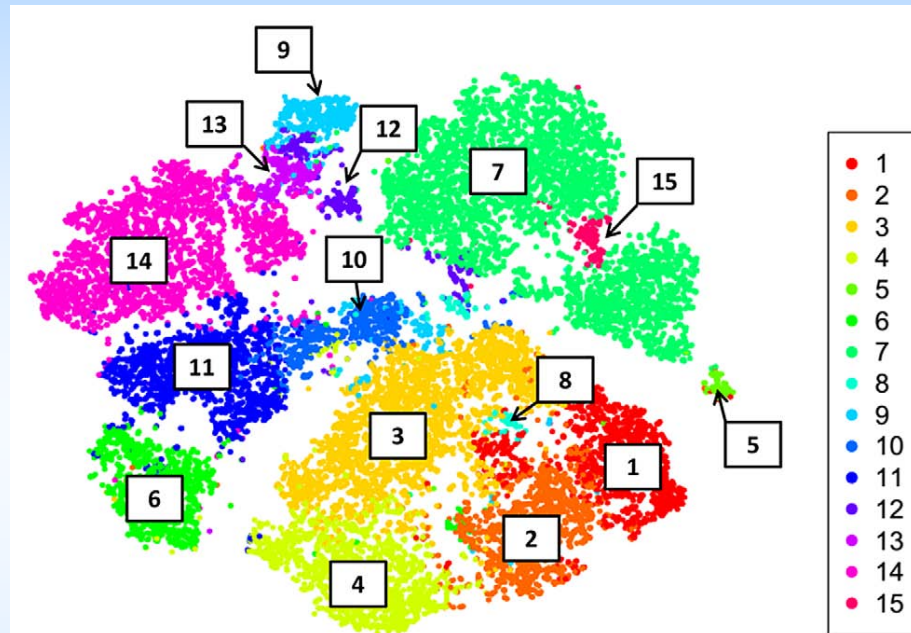


Figure 4. FlowSOM clusters in t-SNE dimensional reduced space. The analysis outputs obtained with the two computational tools were overlaid, and cells displayed as single point in t-SNE map were colored according to FlowSOM metaclusters labeled from 1 to 15. [Color figure can be viewed at wileyonlinelibrary.com]

Lucchesi S, Nolfi E, Pettini E, Pastore G, Fiorino F, Pozzi G, Medaglini D, Ciabattini A.
Computational Analysis of Multiparametric Flow Cytometric Data to Dissect B Cell
Subsets in Vaccine Studies. *Cytometry* (2020) 97A: 259–267

Medicine

- **Goal:** *Individualized* predictions for time course, outcome and disease occurrence in patients
- **Advantage cytometry:** no compartment mixing and easier small entity detection than in genomics or proteomics
- **Problem:** Actual clinical therapy planning is **group** (like *Kaplan-Meier* statistics) and *not individual* patient oriented
- **Improvement:** Use of algorithmic **data pattern** predictions for *individual* patients instead of multidimensional statistics for patient **groups**.

<https://www.classimed.de/classif1.html>

Medicine

- **Status:** „Over the past 20 years, there has been an exponential increase in the number of biomarkers. At the last count, there were **768,259** papers indexed in PubMed.gov directly related to biomarkers. Although any of these papers report clinically useful molecular biomarkers, **embarrassingly** few are currently in clinical use.“

(H.B.Burke, Biomarkers in Cancer **2016** 8:89-99)

Statistics for Group oriented Gene Associations

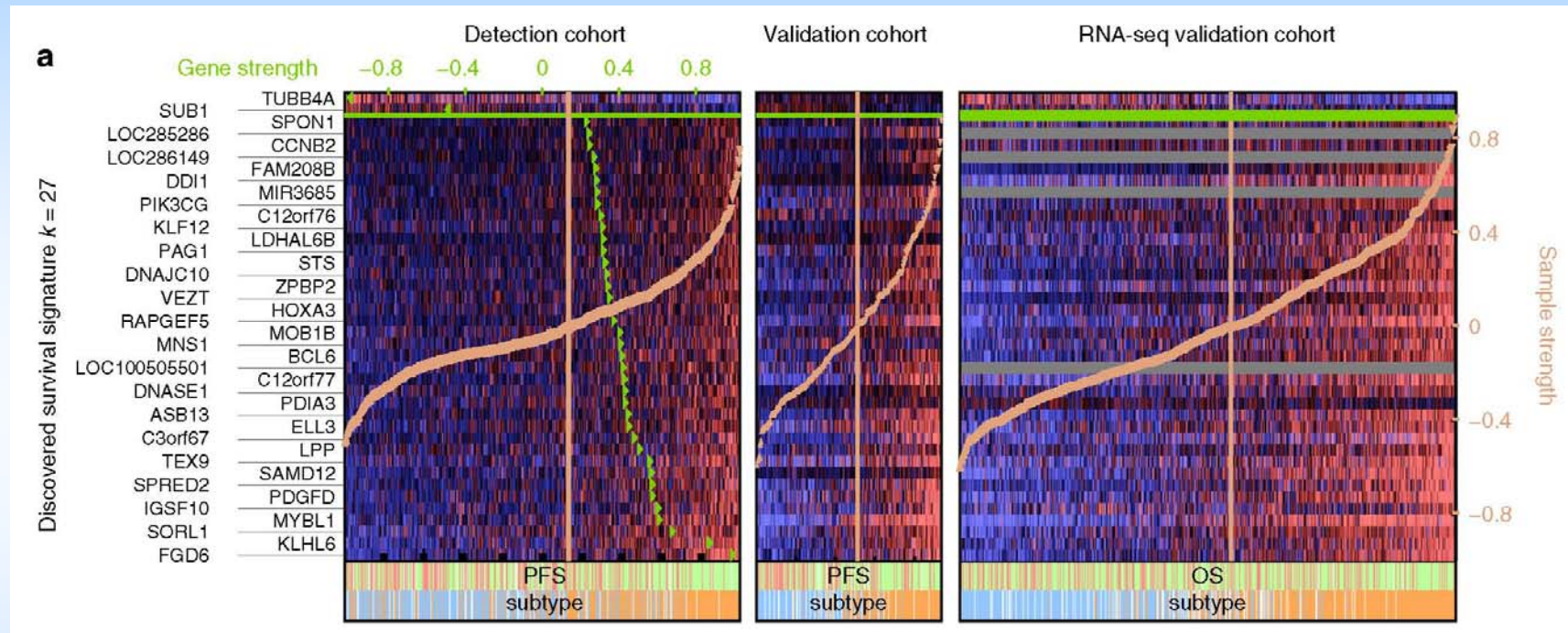
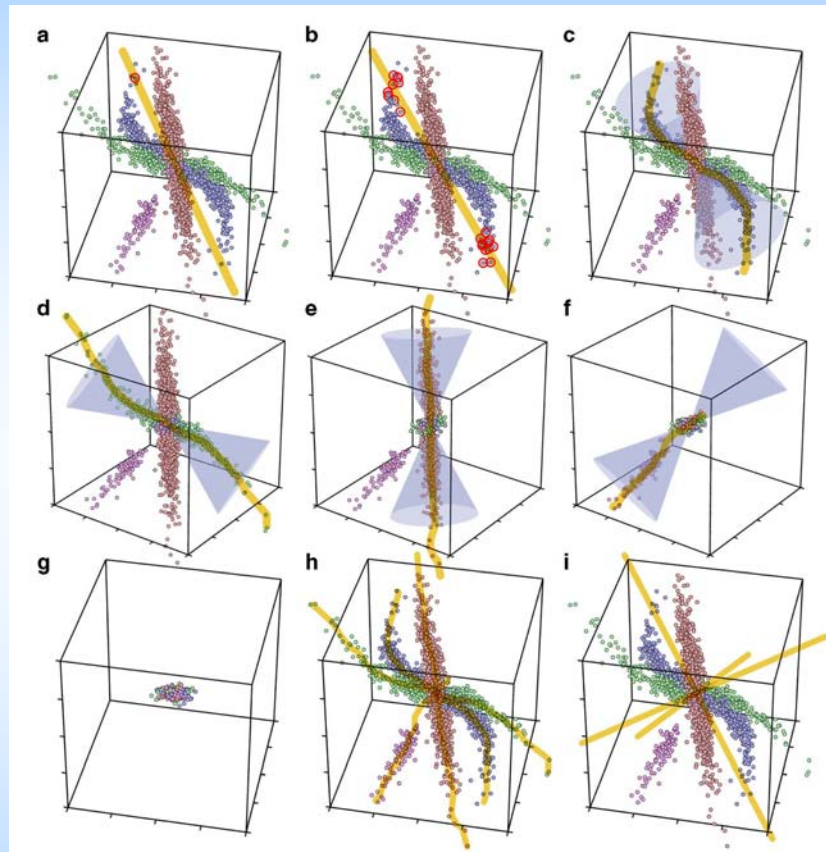


Fig. 5 Discovered survival signatures in **diffuse large B cell lymphoma** (DLBCL, RNA microarrays).

Grau M, Lenz G, Lenz P. Dissection of gene expression datasets into clinically relevant interaction signatures via highdimensional correlation maximization. *Nature Comm* (2019) 10:5417

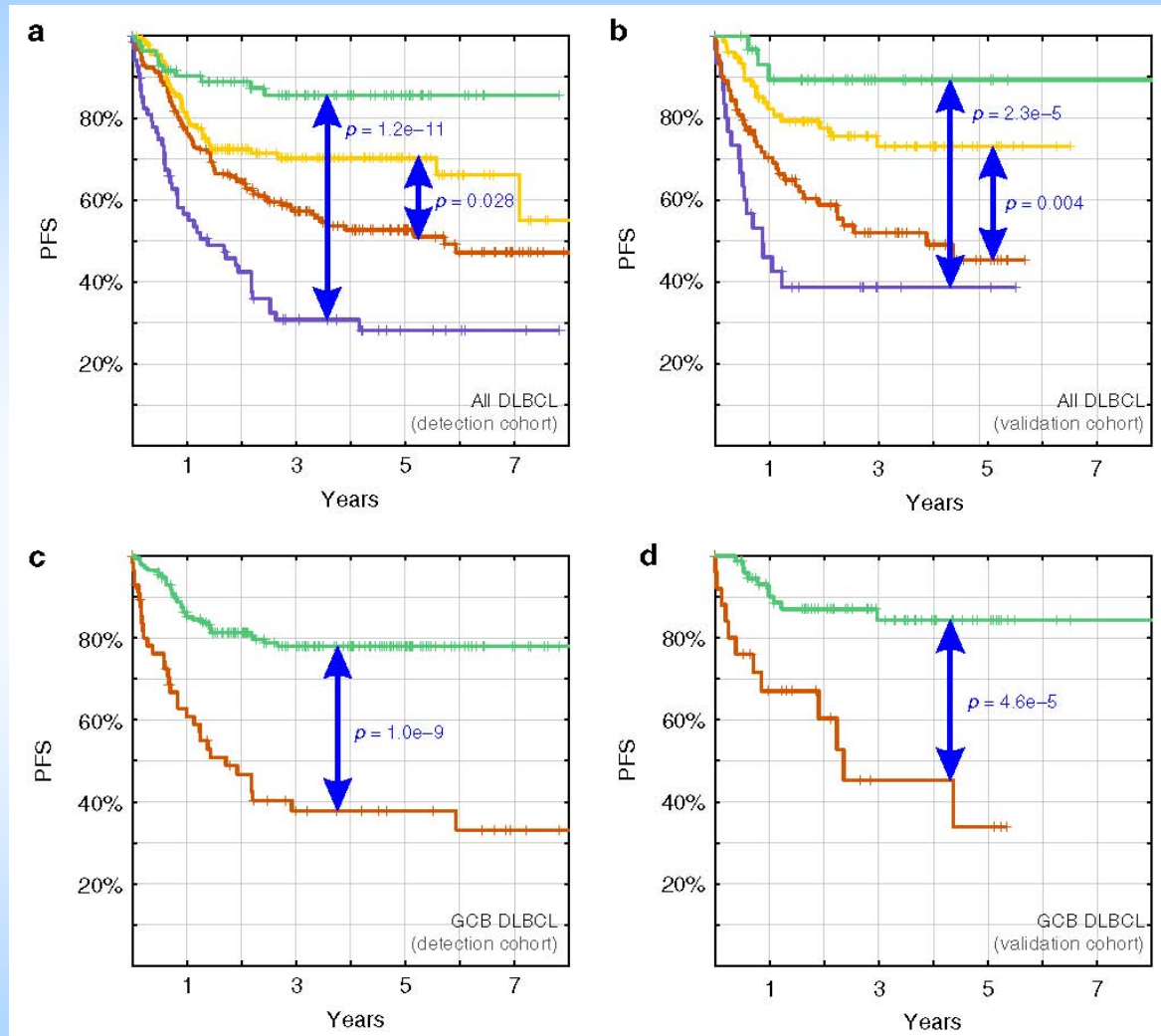
Statistics for **Group** oriented Gene Associations



Diffuse Large B-Cell
Lymphoma (DLBCL)

Fig.1 Concepts of SDCM (Signal Detection by **Correlation Maximization**) illustrated by a 3-dimensional example.

Statistics for Group oriented Gene Associations



Diffuse Large B-Cell Lymphomas (DLBCL)

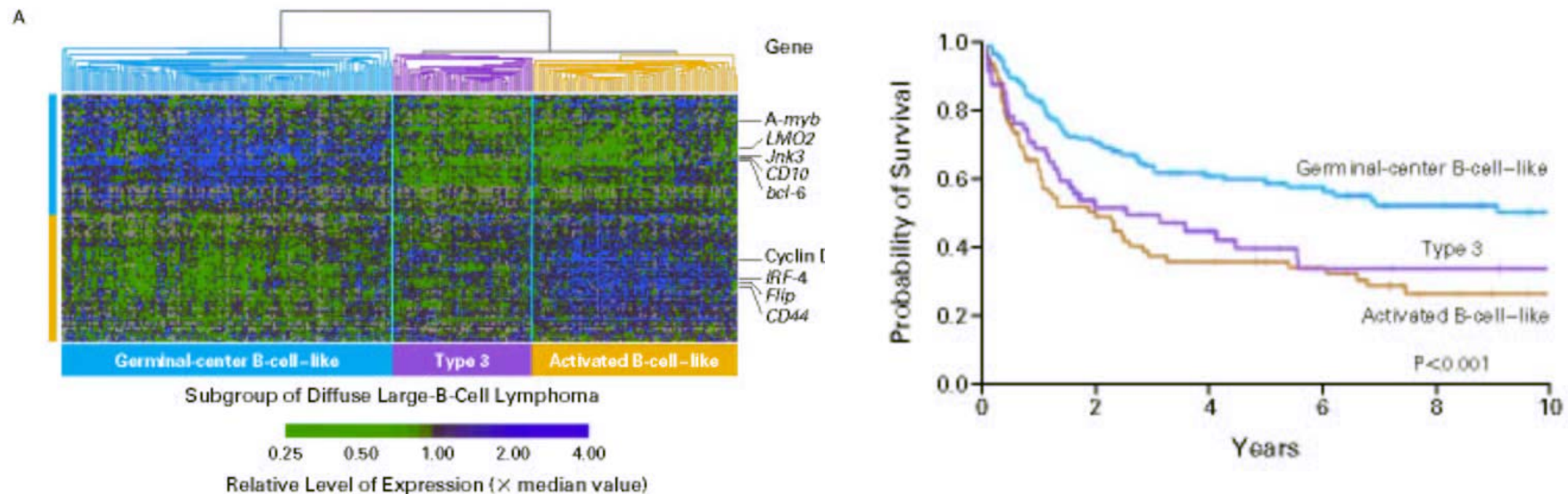
Problems with **Group** Statistics

- only a **fraction** of patients profits from therapy, the others may suffer from therapeutic side effects
- non reactive patients **cannot** be identified in advance with no possibility for **early alternative** therapies
- **patients** are clearly more interested in *individual* disease course and outcome than in **group** statistics
- **gene patterns** of *individual* patients seem more informative for therapy and also for **new hypotheses** development on disease generating mechanisms
- Is it possible to achieve this ? Yes, it is !

Hierarchical Classification for Patient Groups

Diffuse Large B-Cell Lymphoma (DLBCL)

Rosenwald A et al NEJM 346:1937-47 (2002)



hierarchical classification according to Kaplan Meier group statistics

- germinal center B-cell lymphomas
- type 3
- activated B-cell like

“Lymphochip” RNA expression search using 7399 DNA sequences microarray

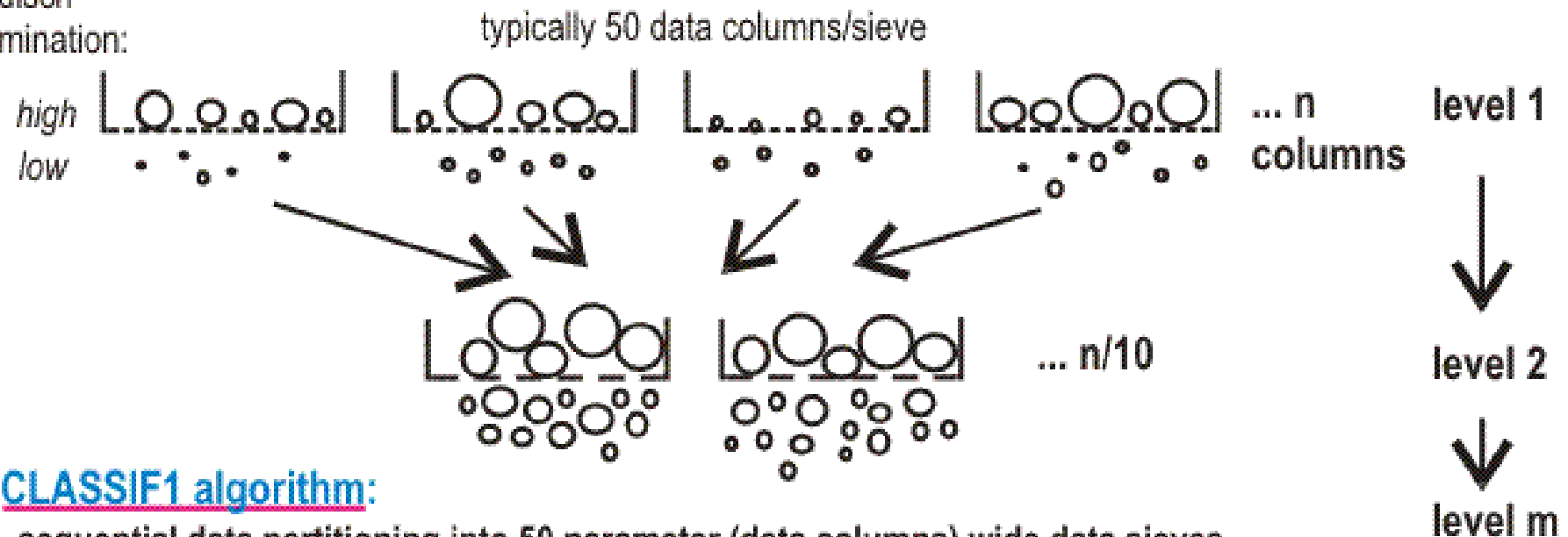
Algorithmic Classification of Individual Patients

by **data pattern** analysis:

- unsupervised
- non hierarchical
- no statistics
- no neuronal network
- no mathematical assumptions
- robust against missing values and outliers
- **standardizable**

Information Enrichment by Data Sieving

discri-
mination:



CLASSIF1 algorithm:

- sequential data partitioning into 50 parameter (data columns) wide data sieves
- select 5 most discriminatory parameters in each sieve by **percentile analysis**
- combine selected parameters, repartition, sieve & repeat procedure until 50 columns or less remain
- classify remaining data columns for **most** discriminatory **triple matrix** data pattern
- algorithm characteristics: **surface data mining** (no models), **unsupervised exhaustive knowledge extraction** to access **unknown knowledge spaces**

GVAL

Data Pattern Classification of Individual Patients

DLBCL Outcome Prediction

data from: NEJM 346:1937-47(2002)

mask pos nr	parameter & UniGene access number	S	NS
5	glutathione synthetase Hs.82327	-0	+
8	MAD mothers against decapentaplegic homolog 4 (Drosophila) Hs.75862	-0	+
10	bone morphogenetic protein 7 (osteogen.prot.1) Hs.170195	-0	+
17	caspase 6, apoptose-related cystein protease Hs.3280	-0	+
20	intercellular adhesion molecule 2 Hs.347326	-0	+
21	chemokine (C-X3-C) receptor 1 Hs.78913	-0	+
23	lymphocyte antigen CD117 Hs.88411	-0	+
1	MAD mothers against decapentaplegic homolog 5 (Drosophila) Hs.37501	0+	-
7	nuclear receptor subfam.3, group C, memeber 3 Hs.75772	0+	-
11	HLA-DPalpha1 Hs.914 *H60848	0+	-
12	HLA-DPalpha1 Hs.914 *H62848	0+	-
13	solute carrier fam.2(facil.glucose transporter) memb.3 Hs.7594	0+	-
15	IFNg inducible protein 30 Hs.14623	0+	-
16	fructose-1,6-biphosphatase 1 Hs.574	0+	-
18	CD9 antigen (p24) Hs.1244	-0	+
19	adenosine kinase Hs.94382	-0	+
2	LC_28024	-0	+
3	DKFZP434F2021 protein Hs.78277	-0	+
4	ESTs Hs.22635	-0	+
6	hypothetical protein MGC4189 Hs.334808	-0	+
9	h.sapiens mRNA, cDNA DKFZp586L 141 Hs.140945	-0	+
22	LC_20218	0+	-
14	hypothetical protein FLJ10116 Hs.79741	0+	-
24	ESTs weakly similar to ALU1_human ALU subfamily J (h.sapiens) Hs.159556	0+	-

val/hoe

- 24 gene data pattern selected from 7399 spot DNA microarray by data sieving
- Individualized data patterns coincide only partially (HLA-DPalpha1) with group patterns

Valet GK, Hoeffkes HG
Data pattern analysis for the individualised pretherapeutic identification of high-risk diffuse large B-cell lymphoma (DLBCL) patients by cytomics
Cytometry (2004) 59A: 232–236

Data Pattern Classification of Individual Patients

Pretherapeutic Risk Assessment in Diffuse Large B-Cell Lymphoma Patients

classify: 7399 array parameters

learning set

unknown test set

clinical outcome	pat. (n)	CLASSIF1 prediction (%)	
		surv	non surv
survival	71	98.6	1.4
non surv.	86	39.5	60.5
neg/pos predval		67.3	98.1

clinical outcome	pat. (n)	CLASSIF1 prediction (%)	
		surv	non surv
survival	29	82.8	17.2
non surv.	47	61.7	38.3
neg/pos predval		45.3	78.3

25-75% percentile thresholds, S1R12P25.BI4

classifiable patients: learn: 157/160 (98.1%), test: 76/80 (95.0%)

Discrimination more informative than Correlation

Individualized Pretherapeutic Risk Assessment in Diffuse Large B-Cell Lymphoma Patients

Data pattern classification of 54630 chip spot parameters (a,b)
versus 105 correlation gene signatures (c,d)

a) chip spot parameter classification				c) signature classific.	
category	patients	surv(%)	nsurv(%)	surv(%)	nsurv(%)
survivor	151	spec 98.7	fpos 1.3	80.1	19.9
non surv	84	fneg 46.4	sens 53.6	54.8	45.2
predval(%)		neg 79.3	pos 95.7	72.5	55.9
b) test set validation				d) test set validation	
survivor	152	92.7	7.3	73.0	27.0
non surv	83	73.5	25.5	60.0	40.0
predval(%)		69.8	66.7	69.0	44.6 <small>gval21</small>

abbreviations: spec=specificity, sens= sensitivity, fp/fn=false positive/
negative, neg/pos= negative/positive predictive values

Sepsis

KE Rudd, SC Johnson, KM Agesa et al. Global, regional, and national **sepsis incidence** and mortality, 1990–2017: analysis for the Global Burden of Disease Study.

Lancet (2020) 395:200-211.

Stern Januar 2020

GEFAHR IM BLUT

Sepsis: Jeder fünfte Todesfall weltweit geht auf das Konto des "unbekannten Killers"

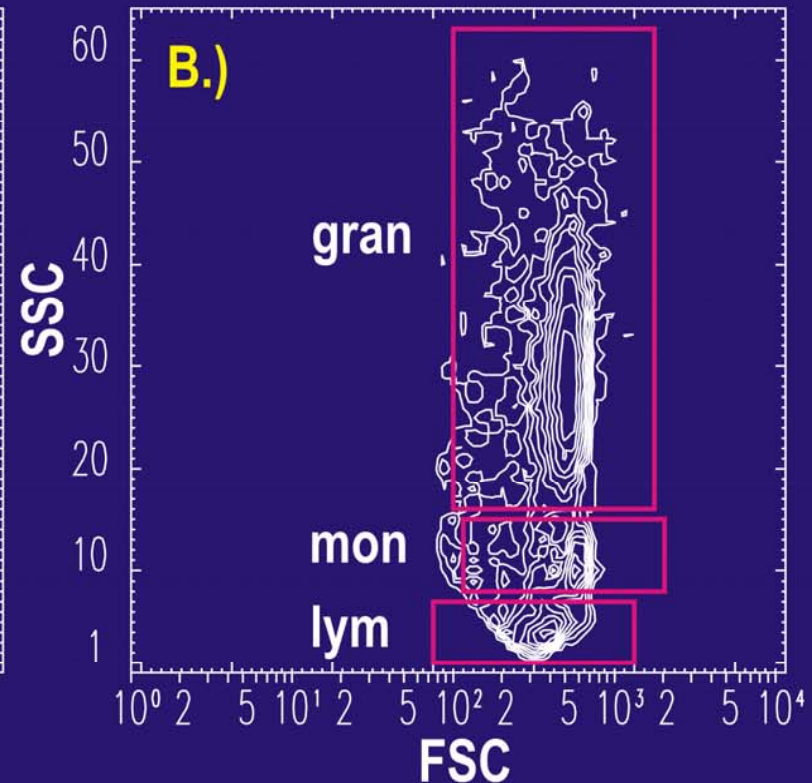
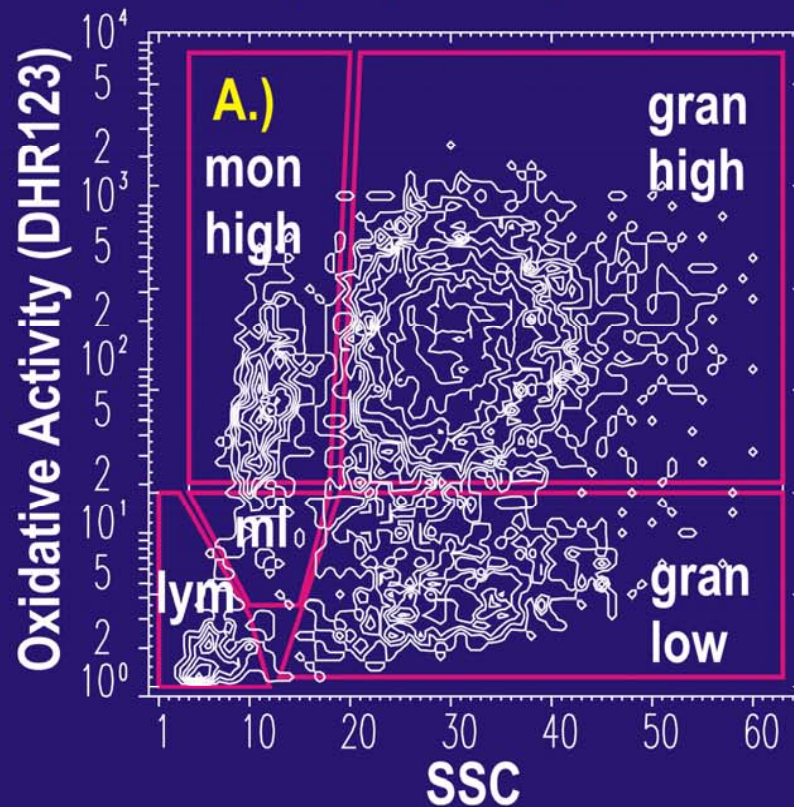


20.01.2020, 20:06 Uhr

Eine Sepsis ist früh erkannt gut zu behandeln, doch verläuft oft tödlich, weil Symptome falsch gedeutet werden. Eine Studie zeigt nun, dass die Krankheit weit mehr Menschen betrifft als bislang angenommen.

Oxidative burst (DHR123) in viable buffy coat blood leukocytes (30min,heparin)

Septic patient, PMA stimulation (100nM), DHR123 (1uM)



Survival prediction (3-15d) for intensive care patients on admission (0d)

#	selected parameters (DHR123)	S0	D0
1	SSC lym (F)	0+	-
2	DHR/FSC high DHR mon (F)	0+	-
3	DHR/FSC all mon (F)	0+	-
4	SSC/FSC all grn (F)	0+	-
5	SSC lym (F+T)	0+	-
6	DHR/FSC lym (F+T)	-0	+
7	% mon of all vital cells (F+T)	-0	+
8	FSC low DHR grn (F+T)	-0	+
9	DHR/FSC low DHR grn (F+T)	-0	+

30-70% percentiles, 9 out of 90 parameters (LSLEARN.B14)

F = FMLP, F+T = FMLP+TNFalpha

gval

Sepsis Risk: **Data Pattern** Analysis for Individual Patients

Nr.	classification category	category abbreviation	class coinc	ref.classification masks
1	survivor	S0	1.00	0 0 0 0 0 0 0 0 + + + + - - - -
2	non surv.	D0	1.00	- - - - + + + +

Nr.	clinical outcome patients: LSLEARN.BI4	CLASSIF1 prediction	class coinc factor	patient classification. masks (.=no value)
2	KE1343 S0	S0	0.67	0 0 + 0 - 0 + 0 +
3	KE1344 S0	S0	0.67	- 0 0 0 - - + 0 0
4	KE1349 S0	S0	0.67	+ - 0 - + - - + -
5	KE1367 S0	S0	0.78	+ 0 0 + 0 - + - +
7	KE1378 S0	S0	0.89	0 + + 0 0 0 0 0 +
8	KE1379 S0	S0	0.67	0 0 - - 0 0 0 + 0
10	KE1386 S0	S0	0.78	+ 0 - 0 + 0 0 + -
12	KE1296 S0	S0	0.67	0 + + - + + + - 0
13	KE1298 S0	S0	0.89	0 + 0 0 0 0 0 - +
14	KE1299 S0	S0	0.78	0 + 0 + - - - + -
16	KE1277 S0	S0	0.78	- - 0 + 0 0 - 0 0
17	KE1292 S0	S0	0.67	- 0 + + - + 0 0 0
18	KE1398 S0	S0	0.56	+ - - 0 + + - + 0
		mask column coincidence (%)		7 7 7 7 6 7 6 6 6 7 7 7 7 9 7 9 2 9
1	KE1334 D0	D0	0.67	- - - 0 - + 0 + 0
6	KE1376 D0	D0	0.89	- - - - - + 0 + +
9	KE1380 D0	D0	0.56	+ 0 - - 0 + + 0 +
11	KE1382 D0	D0	0.78	- - - - - 0 + + 0
15	KE1301 D0	D0	0.56	- + + 0 - + + 0 +
		mask column coincidence (%)		8 6 8 6 8 8 6 6 6 0 0 0 0 0 0 0 0 0

Survival prediction (3-15d) for intensive care patients on **admission** (0d)

- **oxidative burst** in blood leukocytes by flow cytometry
- **stimulation:**
FMLP (100nM),
TNF- α (10ng/ml, 2x10 U/mg prot)
- data pattern **heat map** for **informative** parameters:
(-)=diminished, (0)=unchanged, (+)=increased
LSLEARN.BI4

Valet GK , Roth G, Kellermann W. Risk assessment for intensive care patients by automated classification of flow cytometric data. In: Phagocyte Function, Eds. JP Robinson, GF Babcock, Wiley-Liss Inc, New York 1998, p 289-306.

Sepsis: Data Pattern Analysis for Individual Patients

Prediction of Sepsis Survival on Admission

A. Oxidative Activity (DHR123)

clinical outcome	pat. (n)	CLASSIF1 prediction (%) *	
		surv	non surv
surv	12	100.0	0.0
non surv	5	0.0	100.0
neg/pos predval		100.0	100.0

B. Serine Proteinases (R110)

surv	13	100.0	0.0
non surv	5	20.0	80.0
neg/pos predval		92.9	100.0

* 25-75%, 20-80% percentiles
LSLEARN,KDLEARN.BI4

Challenge

- Cytometrists should address the **systemic potential** of single cell analysis for individualized disease course predictions
- A **human cytome project***) aiming at *predictive medicine by cytomics*, a molecularly standardized *disease classification system* and the establishment of a *periodical system of cells* should be envisaged at the **European level**

*) <https://www.classimed.de/val170.pdf>

Goal: Individualized Predictions instead of Group Statistics

- Core unit cytometrist + clinician
- Retrospective prospective (metaanalysis) data pattern classifications of **all** available patient information (clinical, clinical chemistry, flow cytometry) to elaborate **interlaboratory standardized** classifiers
- Cytometry data standardization:
 - **Intralaboratory** by calibration particles
 - **Interlaboratory** by measurements of a certain number of fresh **blood donor** leukocyte samples, followed by **normalization** of cytometry patient results onto the donor leukocyte means in each laboratory
- **Indistinguishable** blood donor databases by data pattern classification indicate interlaboratory data comparability
- Initial **freedom** of parameter choice for all participating laboratories
- Gradual **selection** of the most discriminatory parameters assures **self focusing** interlaboratory classifier improvement

<https://www.classimed.de/classif1.html#chap6>

30 years DGfZ

- My best **wishes** to the society for efficient future activities and stimulating new ideas for the successful implementation of **patient** oriented applications together with my
- sincere **gratitude** to:
 - all collaborators and collaborating scientists
 - my wife **Hanna** for longterm outstanding work
 - my son **Michael** for essential software developments
 - the **Mildred-Scheel** foundation for generous funding
 - the **Max-Planck Society** as working background for many years

A photograph of a forest path. The path is a narrow, unpaved trail that leads into a dense forest. The trees are tall and slender, with their canopies forming a thick layer of bright green leaves. Sunlight filters through the leaves, creating a dappled pattern of light and shadow on the path and the forest floor. The ground is covered with fallen leaves and small plants. The overall atmosphere is peaceful and natural.

Thanks for Your Attention !

